

***For 60th birthday of BBRC:
DNA replication factors outside S phase***

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DNA replication is one of the most fundamental processes for cell growth. Since the proposal of semi-conservative DNA replication by Watson-Crick and discovery of an enzyme responsible for this process by Kornberg, studies on DNA replication for the past 67 years revealed the involvement of many proteins for regulated execution of DNA replication [1].

Among them, ORC (Origin Recognition Complex), Cdc6, MCM (minichromosome maintenance), and Cdt1 are essential for forming pre-RC, a protein complex required for initiation of DNA replication, on chromatin during G1 phase. Mutations in these genes generally lead to reduced licensing (less pre-RC on the chromatin), resulting in inefficient S phase progression or increased sensitivity to replication stress [2,3].

Extra-replication functions of the pre-RC factors have been indicated. Roles of ORC in transcription silencing were reported shortly after identification of ORC in budding yeast [4,5]. Orc6 subunit of the ORC complex was shown to localize at kinetochores and to the midbody before cytokinesis, and Orc6 depletion led to aberrant nuclear division and mitosis [6,7].

In BBRC 456 (2015) 763–767, Deog Su Hwang's group at Seoul National University reported that Cdc6, an evolutionally conserved replication factor essential for pre-RC formation, is localized to centrosomes during S-G2 phase [8]. The group identified a 56 amino acid segment of Cdc6 which is sufficient for centrosome localization. Similar finding was reported by other group later in the same year [9]. This is an important paper that clearly indicated an important function of Cdc6 in regulation of the centrosome functions. Further studies showed Cdc6 negatively regulates the microtubule-organizing activity of the centrosome by inhibiting the recruitment of pericentriolar material (PCM) proteins to the centrosome [10,11]. Furthermore, Cdc6 was shown to inhibit over-duplication of centrosomes [11,12]. Orc1 was reported to control centrosome copy

number [13].

Orc2 is known to localize to centrosomes [14], and the partner of Cdt1, Geminin, was also reported to associate with centrosomes [15]. The finding by Hwang added Cdc6 to the list of centrosome-associated replication factors. Other groups also reported association of Mcm components to centrosomes [16,17]. Further functional analyses of the centrosome localization of replication factors led to a general conclusion that pre-RC factors negatively regulate centrosome biogenesis.

These findings revealed unexpected roles of pre-RC factors in regulation of centrosome duplication, in addition to its roles in genome duplication. Replication of DNA and duplication of centrosomes are two essential events for cell proliferation, that need to occur “once and only once” during cell cycle in a coordinated manner. pre-RC components play pivotal roles in ensuring ordered and controlled execution of DNA replication and mitosis, firstly by assembly of replication complexes in a proper timing and secondly by defining the timing of centrosome duplication and prevention of its overduplication.

The diverse functions of pre-RC factors in cell cycle progression predict existence of diseases caused by their mutations. Indeed, an autosomal recessive Meier-Gorlin syndrome (MGS) [18-21], an entity of Microcephalic primordial dwarfism, has been shown to be caused by mutations in preRC components including Orc1, Orc4, Orc6, Cdt1 and Cdc6 [22]. In view of the multiple roles of the preRC factors during cell cycle, as described above, mutations in these genes would affect cell proliferation and survival by affecting DNA replication and nuclear/ cell divisions as well as coupling of these processes.

In addition to the roles of pre-RC factors in DNA replication and chromosome separation, a striking defect in the rate of formation of

primary cilia, a modified centriole, was detected in mutants of preRC components. It was also proposed that reduced efficiency in forming cilia could be responsible for clinical features of MGS [23].

Factors participating the processes of DNA replication have been largely identified, and the mechanisms of how they regulate different steps of DNA replication are being elucidated at a rapid rate. However, the functional analyses of replication factors on cellular and an animal level will continue to reveal their unexpected functions. Studies using genetically manipulated model species carrying specific forms of mutations or those bearing a tissue/ organ-specific gene knockout are being awaited to fully understand the biological functions of replication factors.

References

- [1] Masai H, Matsumoto S, You Z, Yoshizawa-Sugata N, Oda M. Eukaryotic chromosome DNA replication: where, when, and how? *Annu Rev Biochem.* 79 (2010) 89-130.
- [2] Ge XQ, Jackson DA, Blow JJ. Dormant origins licensed by excess Mcm2–7 are required for human cells to survive replicative stress. *Genes Dev* 21(2010) 3331–3341.
- [3] Shima N, Alcaraz A, Liachko I, Buske TR, Andrews CA, Munroe RJ, Hartford SA, Tye BK, Schimenti JC. A viable allele of Mcm4 causes chromosome instability and mammary adenocarcinomas in mice. *Nat Genet.* (2007) 93-98.
- [4] Bell SP, Kobayashi R, Stillman B. Yeast origin recognition complex functions in transcription silencing and DNA replication. *Science.* 262 (1993)1844-1849.
- [5] Prasanth SG, Shen Z, Prasanth KV, Stillman B. Human origin recognition complex is essential for HP1 binding to chromatin and heterochromatin organization. *Proc Natl Acad Sci U S A.* 2010 Aug 24;107(34):15093-8.
- [6] Prasanth SG, Prasanth KV, Stillman B. Orc6 involved in DNA replication, chromosome segregation, and cytokinesis. *Science.* 297(2002)1026-1031.
- [7] Bernal JA, Venkitaraman AR. A vertebrate N-end rule degron reveals that Orc6 is required in mitosis for daughter cell abscission. *J Cell Biol.* 192 (2011) 969-978.

- [8] Kim GS, Kang J, Bang SW, Hwang DS. Cdc6 localizes to S- and G2-phase centrosomes in a cell cycle-dependent manner. *Biochem Biophys Res Commun.* 456 (2015) 763-767.
- [9] Kalfalah FM, Berg E, Christensen MO, Linka RM, Dirks WG, Boege F, Mielke C. Spatio-temporal regulation of the human licensing factor Cdc6 in replication and mitosis. *Cell Cycle.* 14 (2015):1704-1715.
- [10] Lee I, Kim GS, Bae JS, Kim J, Rhee K, Hwang DS. The DNA replication protein Cdc6 inhibits the microtubule-organizing activity of the centrosome. *J Biol Chem.* 292 (2017) 16267-16276.
- [11] Xu X, Huang S, Zhang B, Huang F, Chi W, Fu J, Wang G, Li S, Jiang Q, Zhang C. DNA replication licensing factor Cdc6 and Plk4 kinase antagonistically regulate centrosome duplication via Sas-6. *Nat Commun.* 8 (2017)15164.
- [12] Kim GS, Lee I, Kim JH, Hwang DS. The Replication Protein Cdc6 Suppresses Centrosome Over-Duplication in a Manner Independent of Its ATPase Activity. *Mol Cells.* 40 (2017) 925-934.
- [13] Hemerly AS, Prasanth SG, Siddiqui K, Stillman B. Orc1 controls centriole and centrosome copy number in human cells. *Science.* 323 (2009) 789-93.
- [14] Prasanth SG, Prasanth KV, Siddiqui K, Spector DL, Stillman B. Human Orc2 localizes to centrosomes, centromeres and heterochromatin during chromosome inheritance. *EMBO J.* 23(2004) 2651-2663.
- [15] Lu F, Lan R, Zhang H, Jiang Q, Zhang C. Geminin is partially localized to the centrosome and plays a role in proper centrosome duplication. *Biol Cell.* 101(2009) 273-285.
- [16] Stuermer A, Hoehn K, Faul T, Auth T, Brand N, Kneissl M, Pütter V, Grummt F. Mouse pre-replicative complex proteins colocalise and interact with the centrosome. *Eur J Cell Biol.* 86 (2007) 37-50.
- [17] Knockleby J, Lee H. Same partners, different dance: involvement of DNA replication proteins in centrosome regulation. *Cell Cycle.* 9(2010):4487-4491.
- [18] Gorlin RJ, Cervenka J, Moller K, Horrobin M, Witkop CJ Jr. Malformation syndromes. A selected miscellany. *Birth Defects Orig Artic Ser* 11 (1975) 39–50.
- [19] Bongers EM, Opitz JM, Fryer A, Sarda P, Hennekam RC, Hall BD, Superneau DW, Harbison M, Poss A, van Bokhoven H, et al. Meier-Gorlin syndrome: report of eight additional cases and review. *Am J Med Genet* 102 (2001) 115–124.

- [20] Bicknell LS, Walker S, Klingseisen A, Stiff T, Leitch A, Kerzendorfer C, Martin CA, Yeyati P, Al Sanna N, Bober M, Johnson D, Wise C, Jackson AP, O'Driscoll M, Jeggo PA. Mutations in ORC1, encoding the largest subunit of the origin recognition complex, cause microcephalic primordial dwarfism resembling Meier-Gorlin syndrome. *Nat Genet.* 43(2011) 350-355.
- [21] Hossain M, Stillman B. Meier-Gorlin syndrome mutations disrupt an Orc1 CDK inhibitory domain and cause centrosome reduplication. *Genes Dev.* 26 (2012) 1797-810.
- [22] Klingseisen A, Jackson AP. Mechanisms and pathways of growth failure in primordial dwarfism. *Genes Dev.* 25 (2011) 2011-2024.
- [23] Stiff T, Alagoz M, Alcantara D, Outwin E, Brunner HG, Bongers EM, O'Driscoll M, Jeggo PA. Deficiency in origin licensing proteins impairs cilia formation: implications for the aetiology of Meier-Gorlin syndrome. *PLoS Genet.* 9 (2013) e1003360.

Legends to figure

During the G1 phase of the cell cycle, pre-RC factors play essential roles in assembly of a protein complex that is prerequisite for initiation of DNA replication. Pre-RC formation is strictly prohibited once S phase is initiated. Pre-RC factors binds to centrosomes and inhibits overduplication of centrosomes. Its role is generally inhibitory for the centrosome functions, and it may be required for timely and regulated duplication of centrosomes. Pre-RC factors may also inhibit formation of cilia, The two lower panels are cell images from ref. 8 (red, Cdc6 protein), and the upper panel is from ref. 23 (green, acetylated tubulin [entire cilia]; red, γ -tubulin antibodies [the basal body]).

